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Patient Information:
20055, Donor
DOB: [REDACTED]
Sex: M
MR#: [REDACTED]
Patient#: FT-PT8987967

Partner Information:
Not Tested

Physician:
Kuan, JAMES
ATTN: Kuan, JAMES
Las Vegas Sperm Bank
2870 S Maryland Parkway
Ste 250
Lad Vegas, NV 89109
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Laboratory:
Fulgent Therapeutics LLC
CAP#: 8042697
CLIA#: 05D2043189
Laboratory Director:
Dr. Amar Jariwala
Report Date: **Feb 24, 2025**

Accession:
FT-7381585
Test#: FT-TS15095904
Specimen Type: Blood (EDTA)
Collected: Feb 13, 2025

Accession:
N/A

FINAL RESULTS



Carrier for **ONE** genetic condition
Genetic counseling is
recommended.

TEST PERFORMED

Beacon Preconception Carrier Screening - 515 Genes (without X-linked Disorders)

(515 Gene Panel; gene sequencing with deletion and
duplication analysis)

Condition and Gene	Inheritance	20055, Donor	Partner
Very long-chain acyl-CoA dehydrogenase (VLCAD) deficiency ACADVL	AR	⊕ Carrier c.1591C>T (p.Arg531Trp)	N/A

INTERPRETATION:

Notes and Recommendations:

- Based on these results, this individual is positive for a carrier mutation in 1 gene. Carrier screening for the reproductive partner is recommended to accurately assess the risk for any autosomal recessive conditions. A negative result reduces, but does not eliminate, the chance to be a carrier for any condition included in this screen. Please see the supplemental table for details.
- Testing for copy number changes in the SMN1 gene was performed to screen for the carrier status of Spinal Muscular Atrophy. The results for this individual are within the normal range for non-carriers. See Limitations section for more information.
- This carrier screening test does not screen for all possible genetic conditions, nor for all possible mutations in every gene tested. This report does not include variants of uncertain significance; only variants classified as pathogenic or likely pathogenic at the time of testing, and considered relevant for reproductive carrier screening, are reported. Please see the gene specific notes for details. Please note that the classification of variants can change over time.
- Patients may wish to discuss any carrier results with blood relatives, as there is an increased chance that they are also carriers. These results should be interpreted in the context of this individual's clinical findings, biochemical profile, and family history.
- X-linked genes are not routinely analyzed for male carrier screening tests. Gene specific notes and limitations may be present. See below.
- Genetic counseling is recommended. Available genetic counselors and additional resources can be found at the National Society of Genetic Counselors (NSGC; <https://www.nsgc.org>)

Patient: 20055, Donor; Sex: M;
DOB: [REDACTED] **MR#:** [REDACTED]

Accession#: FT-7381585; FD Patient#: FT-PT8987967;
DocID: FT-TS15095904AA; **PAGE 1 of 5**



VERY LONG-CHAIN ACYL-COA DEHYDROGENASE (VLCAD) DEFICIENCY

Patient	20055, Donor	Partner
Result	⊕ Carrier	N/A
Variant Details	ACADVL (NM_000018.4) c.1591C>T (p.Arg531Trp)	N/A

What is Very long-chain acyl-CoA dehydrogenase (VLCAD) deficiency?

VLCAD deficiency is a condition in which the body is unable to properly breakdown very long-chain fatty acids into energy, particularly during periods of fasting. There are three forms of VLCAD deficiency: a severe, early-onset form; a hepatic (liver) or hypoketotic hypoglycemic form; and a later-onset episodic myopathic form. Signs and symptoms of the severe, early-onset form occur in the first few months of life and include cardiomyopathy, pericardial effusion, heart arrhythmias, low muscle tone, enlarged liver, and intermittent hypoglycemia. People affected by the hepatic or hypoketotic hypoglycemic form typically develop symptoms during early childhood. These features may include hypoketotic hypoglycemia and an enlarged liver (without cardiomyopathy). Signs and symptoms of the later-onset episodic myopathic form may include intermittent rhabdomyolysis, muscle cramps, muscle pain, and exercise intolerance. It is the most common form of VLCAD deficiency.

What is my risk of having an affected child?

VLCAD is inherited in an autosomal recessive manner. When both parents are carriers for the same condition, there is a 25% (1/4) risk to have an affected child.

What kind of medical management is available?

Management of VLCAD deficiency depends on the form of the condition and the specific signs and symptoms present. People affected by the severe forms of the condition are typically placed on a low-fat, high-carbohydrate diet with frequent meals. Periods of rhabdomyolysis may be treated with hydration and alkalization of the urine to protect kidney function and prevent acute kidney failure. Affected people are generally advised to avoid fasting, dehydration, and a high-fat diet. The prognosis for people with VLCAD deficiency is hard to predict and can vary based on the severity of the condition. For people with the severe, early-onset form, cardiomyopathy and arrhythmias can be lethal. However, heart function can be improved with early treatment and specific diet modifications. Normal intellectual outcome has been reported in people with this form. For people with the hepatic or hypoketotic hypoglycemic form, hypoketotic hypoglycemia can cause a loss of consciousness or seizures. People with the later-onset episodic myopathic form may only have muscle-related symptoms. Some people with this form may not be diagnosed until adulthood. Since the later-onset form can have vague or intermittent symptoms, it is possible that some people may have no symptoms during their lifetime.

What mutation was detected?

The detected heterozygous variant was NM_000018.4:c.1591C>T (p.Arg531Trp). This variant has been reported in the presumed or confirmed compound heterozygous state in at least four unrelated individuals with very long chain acyl-CoA dehydrogenase deficiency (PubMed: [21932095](#), [26385305](#), [31031081](#), [35281659](#), Invitae personal communication). Additionally, another variant at this position in the gene (p.Arg531Leu) has been associated with VLCAD deficiency, suggesting that a change at this position adversely affects protein structure and/or function and is potentially disease-causing (PubMed: [27209629](#)). This variant has been identified by this laboratory in the compound heterozygous state in 1 individual with symptoms consistent with an ACADVL-related condition. The laboratory classifies this variant as likely pathogenic.



GENES TESTED:

Beacon Preconception Carrier Screening - 515 Genes (without X-linked Disorders) - 515 Genes

This analysis was run using the Beacon Preconception Carrier Screening - 515 Genes (without X-linked Disorders) gene list. 515 genes were tested with 99.7% of targets sequenced at >20x coverage. For more gene-specific information and assistance with residual risk calculation, see the SUPPLEMENTAL TABLE.

AAAS, ABCA12, ABCA3, ABCA4, ABCB11, ABCB4, ABCC2, ABCC8, ACAD9, ACADM, ACADVL, ACAT1, ACOX1, ACSF3, ADA, ADAMTS2, ADAMTSL4, ADGRG1, ADGRV1, AGA, AGL, AGPS, AGXT, AHI1, AIPL1, AIRE, ALDH3A2, ALDH7A1, ALDOB, ALG1, ALG6, ALMS1, ALPL, AMN, AMT, ANO10, AP1S1, AQP2, ARG1, ARL6, ARSA, ARSB, ASL, ASNS, ASPA, ASS1, ATM, ATP6V1B1, ATP7B, ATP8B1, BBS1, BBS10, BBS12, BBS2, BBS4, BBS5, BBS7, BBS9, BCKDHA, BCKDHB, BCS1L, BLM, BLOC1S3, BLOC1S6, BMP1, BRIP1, BSND, CAD, CANT1, CAPN3, CASQ2, CBS, CC2D1A, CC2D2A, CCDC103, CCDC39, CCDC88C, CD3D, CD3E, CD40, CD59, CDH23, CEP152, CEP290, CERKL, CFTR, CHAT, CHRNE, CHRNA, CIITA, CLCN1, CLN3, CLN5, CLN6, CLN8, CLRN1, CNGB3, COL11A2, COL17A1, COL27A1, COL4A3, COL4A4, COL7A1, COX15, CPS1, CPT1A, CPT2, CRB1, CRTAP, CRYL1, CTNS, CTSA, CTSC, CTSD, CTSK, CYBA, CYP11A1, CYP11B1, CYP11B2, CYP17A1, CYP19A1, CYP1B1, CYP21A2, CYP27A1, CYP27B1, CYP7B1, DBT, DCAF17, DCLRE1C, DDX11, DGAT1, DGUOK, DHCR7, DHDDS, DLD, DLL3, DNAH11, DNAH5, DNAI1, DNAI2, DNMT3B, DOK7, DUOX2, DYNC2H1, DYSF, EIF2AK3, EIF2B1, EIF2B2, EIF2B3, EIF2B4, EIF2B5, ELP1, EPG5, ERCC2, ERCC6, ERCC8, ESCO2, ETFA, ETFB, ETFDH, ETHE1, EVC, EVC2, EXOSC3, EYS, FAH, FAM161A, FANCA, FANCC, FANCD2, FANCE, FANCG, FANCI, FANCL, FBP1, FBXO7, FH, FKBP10, FKBP, FKTN, FMO3, FOXN1, FOXRED1, FRAS1, FREM2, FUCA1, G6PC, G6PC3, GAA, GALT, GALE, GALK1, GALNS, GALNT3, GALT, GAMT, GATM, GBA, GBE1, GCDH, GCH1, GDF5, GFM1, GHR, GJB2, GJB6, GJB1, GLDC, GLE1, GNE, GNPAT, GNPTAB, GNPTG, GNS, GORAB, GRHR, GRIP1, GSS, GUCY2D, GUSB, HADH, HADHA, HADHB, HAMP, HAX1, HBA1, HBA2, HBB, HEXA, HEXB, HGSNAT, HJV, HLCS, HMGCL, HMOX1, HOGA1, HPD, HPS1, HPS3, HPS4, HPS5, HPS6, HSD17B3, HSD17B4, HSD3B2, HYAL1, HYLS1, IDUA, IGHMBP2, IKBK, IL7R, INVS, ITGA6, ITGB3, ITGB4, IVD, JAK3, KCNJ1, KCNJ11, LAMA2, LAMA3, LAMB3, LAMC2, LARGE1, LCA5, LDLR, LDLRAP1, LHX3, LIFR, LIG4, LIPA, LMBRD1, LOXHD1, LPL, LRAT, LRP2, LRPPRC, LYST, MAK, MAN2B1, MANBA, MCEE, MCOLN1, MCPH1, MECP, MED17, MESF2, MFSDB, MKKS, MKS1, MLC1, MLYCD, MMAA, MMAB, MMACHC, MMADHC, MOCS1, MOCS2, MPI, MPL, MPV17, MRE11, MTHFR, MTR, MTRR, MTPP, MUSK, MUT, MVK, MYO15A, MYO7A, NAGA, NAGLU, NAGS, NBN, NCF2, NDRG1, NDUFAF2, NDUFAF5, NDUFS4, NDUFS6, NDUFS7, NDUFV1, NEB, NEU1, NGLY1, NPC1, NPC2, NPHP1, NPHS1, NPHS2, NR2E3, NSMCE3, NTRK1, OAT, OCA2, OPA3, OSTM1, OTOA, OTOF, P3H1, PAH, PANK2, PC, PCBD1, PCCA, PCCB, PCDH15, PCNT, PDHB, PEPD, PET100, PEX1, PEX10, PEX12, PEX13, PEX16, PEX2, PEX26, PEX5, PEX6, PEX7, PFKM, PGM3, PHGDH, PHKB, PHKG2, PHYH, PIGN, PJK, PKHD1, PLA2G6, PLEKHG5, PLOD1, PMM2, PNPO, POLG, POLH, POMGNT1, POMT1, POMT2, POR, POU1F1, PPT1, PRCD, PRDM5, PRF1, PROP1, PSAP, PTPRC, PTS, PUS1, PYGM, QDPR, RAB23, RAG1, RAG2, RAPSN, RARS2, RDH12, RLBP1, RMRP, RNASEH2A, RNASEH2B, RNASEH2C, RPE65, RPGRIP1L, RTEL1, RXYLT1, RYR1, SACS, SAMD9, SAMHD1, SCO2, SEC23B, SEPSecs, SGCA, SGCB, SGCD, SGCG, SGSH, SKIV2L, SLC12A1, SLC12A3, SLC12A6, SLC17A5, SLC19A2, SLC19A3, SLC1A4, SLC22A5, SLC25A13, SLC25A15, SLC25A20, SLC26A2, SLC26A3, SLC26A4, SLC27A4, SLC35A3, SLC37A4, SLC38A8, SLC39A4, SLC45A2, SLC4A11, SLC5A5, SLC7A7, SMARCA1, SMN1, SMPD1, SNAP29, SPG11, SPR, SRD5A2, ST3GAL5, STAR, STX11, STXBP2, SUMF1, SUOX, SURF1, SYNE4, TANGO2, TAT, TBCD, TBCE, TCIRG1, TCN2, TECPR2, TERT, TF, TFR2, TG, TGM1, TH, TK2, TMC1, TMEM216, TMEM67, TMPPRS3, TPO, TPP1, TREX1, TRIM32, TRIM37, TRMU, TSEN54, TSFM, TSHB, TSHR, TTC37, TTPA, TULP1, TYMP, TYR, TYRP1, UBR1, UNC13D, USH1C, USH2A, VDR, VLDLR, VPS11, VPS13A, VPS13B, VPS45, VPS53, VRK1, VSX2, WISP3, WNT10A, WRN, XPA, XPC, ZBTB24, ZFYVE26, ZNF469

METHODS:

Genomic DNA was isolated from the submitted specimen indicated above (if cellular material was submitted). DNA was barcoded, and enriched for the coding exons of targeted genes using hybrid capture technology. Prepared DNA libraries were then sequenced using a Next Generation Sequencing technology. Following alignment to the human genome reference sequence (assembly GRCh37), variants were detected in regions of at least 10x coverage. For this specimen, 99.74% and 99.68% of coding regions and splicing junctions of genes listed had been sequenced with coverage of at least 10x and 20x, respectively, by NGS or by Sanger sequencing. The remaining regions did not have 10x coverage, and were not evaluated. Variants were interpreted manually using locus specific databases, literature searches, and other molecular biological principles. To minimize false positive results, any variants that do not meet internal quality standards are confirmed by Sanger sequencing. Variants classified as pathogenic, likely pathogenic, or risk allele which are located in the coding regions and nearby intronic regions (+/- 20bp) of the genes listed above are reported. Variants outside these intervals may be reported but are typically not guaranteed. When a single pathogenic or likely pathogenic variant is identified in a clinically relevant gene with autosomal recessive inheritance, the laboratory will attempt to ensure 100% coverage of coding sequences either through NGS or Sanger sequencing technologies ("fill-in"). All genes listed were evaluated for large deletions and/or duplications. However, single exon deletions or duplications will not be detected in this assay, nor will copy number alterations in regions of genes with significant pseudogenes. Putative deletions or duplications are analyzed using Fulgent Germline proprietary pipeline for this specimen. Bioinformatics: The FPLMv2.0 pipeline was used to analyze this specimen.

LIMITATIONS:

General Limitations

These test results and variant interpretation are based on the proper identification of the submitted specimen, accuracy of any stated familial relationships, and use of the correct human reference sequences at the queried loci. In very rare instances, errors may result due to mix-up or co-mingling of specimens. Positive results do not imply that there are no other contributors, genetic or



otherwise, to future pregnancies, and negative results do not rule out the genetic risk to a pregnancy. Official gene names change over time. Fulgent uses the most up to date gene names based on HUGO Gene Nomenclature Committee (<https://www.genenames.org>) recommendations. If the gene name on report does not match that of ordered gene, please contact the laboratory and details can be provided. Result interpretation is based on the available clinical and family history information for this individual, collected published information, and Alamut annotation available at the time of reporting. This assay is not designed or validated for the detection of low-level mosaicism or somatic mutations. This assay will not detect certain types of genomic aberrations such as translocations, inversions, or repeat expansions other than specified genes. DNA alterations in regulatory regions or deep intronic regions (greater than 20bp from an exon) may not be detected by this test. Unless otherwise indicated, no additional assays have been performed to evaluate genetic changes in this specimen. There are technical limitations on the ability of DNA sequencing to detect small insertions and deletions. Our laboratory uses a sensitive detection algorithm, however these types of alterations are not detected as reliably as single nucleotide variants. Rarely, due to systematic chemical, computational, or human error, DNA variants may be missed. Although next generation sequencing technologies and our bioinformatics analysis significantly reduce the confounding contribution of pseudogene sequences or other highly-homologous sequences, sometimes these may still interfere with the technical ability of the assay to identify pathogenic alterations in both sequencing and deletion/duplication analyses. Deletion/duplication analysis can identify alterations of genomic regions which include one whole gene (buccal swab specimens and whole blood specimens) and are two or more contiguous exons in size (whole blood specimens only); single exon deletions or duplications may occasionally be identified, but are not routinely detected by this test. When novel DNA duplications are identified, it is not possible to discern the genomic location or orientation of the duplicated segment, hence the effect of the duplication cannot be predicted. Where deletions are detected, it is not always possible to determine whether the predicted product will remain in-frame or not. Unless otherwise indicated, deletion/duplication analysis has not been performed in regions that have been sequenced by Sanger.

Gene Specific Notes and Limitations

ALG1: Due to the interference by highly homologous regions, our current testing method has less sensitivity to detect variants in exons 6-13 of the ALG1 gene (NM_019109.4). **CEP290:** Copy number analysis for exons 8-13 and exons 39-42 may have reduced sensitivity in the CEP290 gene. Confirmation of these exons are limited to individuals with a positive personal history of CEP290-related conditions and/or individuals carrying a pathogenic/likely pathogenic sequence variant. **CFTR:** Analysis of the intron 8 polymorphic region (e.g. IVS8-5T allele) is only performed if the p.Arg117His (R117H) mutation is detected. Single exon deletion/duplication analysis is limited to deletions of previously reported exons: 1, 2, 3, 11, 19, 20, 21. Analysis of the intron 8 polymorphic region (e.g. IVS8-5T allele) is only performed if the p.Arg117His (R117H) mutation is detected. Single exon deletion/duplication analysis is limited to deletions of previously reported exons: 1, 2, 3, 11, 19, 20, 21. CFTR variants primarily associated with CFTR-related isolated congenital bilateral absence of the vas deferens and CFTR-related pancreatitis are not included in this analysis. CFTR variants with insufficient evidence of being cystic fibrosis mutations will not be reported either. **CRYL1:** As mutations in the CRYL1 gene are not known to be associated with any clinical condition, sequence variants in this gene are not analyzed. However, to increase copy number detection sensitivity for large deletions, this gene was evaluated for copy number variation. **CYP11B1:** The current testing method is not able to reliably detect certain pathogenic variants in this gene due to the interference by highly homologous regions. This analysis is not designed to detect or rule-out copy-neutral chimeric CYP11B1/CYP11B2 gene. **CYP11B2:** The current testing method is not able to reliably detect certain pathogenic variants in this gene due to the interference by highly homologous regions. This analysis is not designed to detect or rule-out copy-neutral chimeric CYP11B1/CYP11B2 gene. **CYP21A2:** Significant pseudogene interference and/or reciprocal exchanges between the CYP21A2 gene and its pseudogene, CYP21A1P, have been known to occur and may impact results. As such, the relevance of variants reported in this gene must be interpreted clinically in the context of the clinical findings, biochemical profile, and family history of each patient. LR-PCR is not routinely ordered for NM_000500.9:c.955C>T (p.Gln319Ter). Individuals with c.955C>T (p.Gln319Ter) will be reported as a Possible Carrier indicating that the precise nature of the variant has not been determined by LR-PCR and that the variant may occur in the CYP21A2 wild-type gene or in the CYP21A1P pseudogene. The confirmation test is recommended if the second reproductive partner is tested positive for variants associated with classic CAH. **DDX11:** Due to the interference by highly homologous regions, our current testing method has less sensitivity to detect variants in the DDX11 gene. **DUOX2:** The current testing method is not able to reliably detect variants in exons 6-8 of the DUOX2 gene (NM_014080.5) due to significant interference by the highly homologous gene, DUOX1. **FANCD2:** Due to pseudogene interference, copy-number-variants within exon 14-17 of the FANCD2 gene (NM_033084.4) are not evaluated and detection of single-nucleotide variants and small insertions/deletions in this region is not guaranteed. **GALT:** In general, the D2 "Duarte" allele is not reported if detected, but can be reported upon request. While this allele can cause positive newborn screening results, it is not known to cause clinical symptoms in any state. See GeneReviews for more information: <https://www.ncbi.nlm.nih.gov/books/NBK1518/> **GBA:** Significant pseudogene interference and/or reciprocal exchanges between the GBA gene and its pseudogene, GBAP1, have been known to occur and may impact results. As such, the relevance of variants reported in this gene must be interpreted clinically in the context of this individual's clinical findings, biochemical profile, and family history. The current testing method cannot detect copy-neutral rearrangements between the pseudogene and the functional gene, which have been reported in very rare cases of Gaucher disease (PubMed: 21704274). **HBA1:** Significant interference from highly homologous regions in exons 1-2 of the HBA1 gene has



been recognized to occur, potentially impeding the assay's technical capability to detect pathogenic alterations during sequencing analyses. **HBA2:** Significant interference from highly homologous regions in exons 1-2 of the HBA2 gene has been recognized to occur, potentially impeding the assay's technical capability to detect pathogenic alterations during sequencing analyses. **HSD17B4:** Copy number analysis for exons 4-6 may have reduced sensitivity in the HSD17B4 gene. Confirmation of these exons are limited to individuals with a positive personal history of D-bifunctional protein deficiency and Perrault syndrome and/or individuals carrying a pathogenic/likely pathogenic sequence variant. **LMBRD1:** Copy number analysis for exons 9-12 may have reduced sensitivity in the LMBRD1 gene. Confirmation of these exons are limited to individuals with a positive personal history of combined methylmalonic aciduria and homocystinuria and/or individuals carrying a pathogenic/likely pathogenic sequence variant. **MTHFR:** As recommended by ACMG, the two common polymorphisms in the MTHFR gene - c.1286A>C (p.Glu429Ala, also known as c.1298A>C) and c.665C>T (p.Ala222Val, also known as c.677C>T) - are not reported in this test due to lack of sufficient clinical utility to merit testing (PubMed: [23288205](#)). **NEB:** This gene contains a 32-kb triplicate region (exons 82-105) which is not amenable to sequencing and deletion/duplication analysis. **NPHS2:** If detected, the variant NM_014625.3:c.686G>A (p.Arg229Gln) will not be reported as this variant is not significantly associated with disease when homozygous or in the compound heterozygous state with variants in exons 1-6 of NPHS2. **OTOA:** Due to pseudogene interference, our current testing method is not able to reliably detect variants in exons 20-28 (NM_144672.3) in the OTOA gene. **SMN1:** The current testing method detects sequencing variants in exon 7 and copy number variations in exons 7-8 of the SMN1 gene (NM_022874.2). Sequencing and deletion/duplication analysis are not performed on any other region in this gene. About 5%-8% of the population have two copies of SMN1 on a single chromosome and a deletion on the other chromosome, known as a [2+0] configuration (PubMed: [20301526](#)). The current testing method cannot directly detect carriers with a [2+0] SMN1 configuration but can detect linkage between the silent carrier allele and certain population-specific single nucleotide changes. As a result, a negative result for carrier testing greatly reduces but does not eliminate the chance that a person is a carrier. Only abnormal results will be reported. **TERT:** The TERT promoter region is analyzed for both sequencing and copy number variants. **TYR:** Due to the interference by highly homologous regions, our current testing method has less sensitivity to detect variants in exons 4-5 of the TYR gene (NM_000372.5). **WRN:** Due to the interference by highly homologous regions within the WRN gene, our current testing method has less sensitivity to detect variants in exons 10-11 of WRN (NM_000553.6).

SIGNATURE:



Yan Meng, Ph.D., CGMB, FACMG on 2/24/2025
Laboratory Director, Fulgent

DISCLAIMER:

This test was developed and its performance characteristics determined by Fulgent Therapeutics LLC CAP #8042697 CLIA #05D2043189; 4399 Santa Anita Ave., El Monte, CA, 91731. It has not been cleared or approved by the FDA. The laboratory is regulated under CLIA as qualified to perform high-complexity testing. This test is used for clinical purposes. It should not be regarded as investigational or for research. Since genetic variation, as well as systematic and technical factors, can affect the accuracy of testing, the results of testing should always be interpreted in the context of clinical and familial data. For assistance with interpretation of these results, healthcare professionals may contact us directly at [626-350-0537](tel:626-350-0537) or by email at info@fulgentgenetics.com. It is recommended that patients receive appropriate genetic counseling to explain the implications of the test result, including its residual risks, uncertainties and reproductive or medical options.

To view the supplemental table describing the carrier frequencies, detection rates, and residual risks associated with the genes tested on any Beacon panel, please visit the following link:

[Beacon Expanded Carrier Screening Supplemental Table](#)





Patient Information:

20055, Donor

DOB: [REDACTED]

Sex: M

MR#:

Patient#: FT-PT8987967

Partner Information:

Not Tested

Physician:

Kuan, JAMES

ATTN: Kuan, JAMES

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CLIA#: 05D2043189

Laboratory Director:

Dr. Amar Jariwala

Report Date: **Aug 17, 2026**

Accession:

FT-7381585

Test#: FT-TS15658814

Specimen Type: Blood (EDTA)

Collected: Feb 13, 2025

Accession:

N/A

FINAL RESULTS



Carrier for **ONE** genetic condition

TEST PERFORMED

Custom Beacon Preconception Carrier Screening Panel

(2 Gene Panel: *BTB* and *F2*; gene sequencing with deletion and duplication analysis)

Condition and Gene	Inheritance	20055, Donor	Partner
Biotinidase deficiency <i>BTB</i>	AR	+ Carrier c.1270G>C (p.Asp424His)	N/A

INTERPRETATION:

Notes and Recommendations:

- Based on these results, this individual is positive for a carrier mutation in 1 gene. Carrier screening for the reproductive partner is recommended to accurately assess the risk for any autosomal recessive conditions. A negative result reduces, but does not eliminate, the chance to be a carrier for any condition included in this screen. Please see the supplemental table for details.
- This carrier screening test does not screen for all possible genetic conditions, nor for all possible mutations in every gene tested. This report does not include variants of uncertain significance; only variants classified as pathogenic or likely pathogenic at the time of testing, and considered relevant for reproductive carrier screening, are reported. Please see the gene specific notes for details. Please note that the classification of variants can change over time.
- Patients may wish to discuss any carrier results with blood relatives, as there is an increased chance that they are also carriers. These results should be interpreted in the context of this individual's clinical findings, biochemical profile, and family history.
- Gene specific notes and limitations may be present. See below.
- Genetic counseling is recommended. Available genetic counselors and additional resources can be found at the National Society of Genetic Counselors (NSGC; <https://www.nsgc.org>).



BIOTINIDASE DEFICIENCY

Patient	20055, Donor	Partner
Result	 Carrier	N/A
Variant Details	BTD (NM_001370658.1) c.1270G>C (p.Asp424His)	N/A

What is Biotinidase deficiency?

Biotinidase deficiency is a disorder in which the body is unable to recycle the vitamin biotin. If this condition is not recognized and treated, its signs and symptoms typically appear within the first few months of life, although it can also become apparent later in childhood. Profound biotinidase deficiency, the more severe form of the condition, can cause seizures, hypotonia, breathing problems, hearing and vision loss, ataxia, skin rashes, alopecia, and a fungal infection called candidiasis. Affected children also have delayed development. Partial biotinidase deficiency is a milder form of this condition. Without treatment, affected children may experience hypotonia, skin rashes, and hair loss, but these problems may appear only during illness, infection, or other times of stress.

What is my risk of having an affected child?

Biotinidase deficiency is inherited in an autosomal recessive manner. If the patient and the partner are both carriers, the risk for an affected child is 1 in 4 (25%).

What kind of medical management is available?

Biotinidase deficiency is detected on newborn screening. Lifelong treatment with vitamin supplementation can prevent these complications from occurring or improve them if they have already developed. Without treatment, affected children may experience hypotonia, skin rashes, and hair loss, but these problems may appear only during illness, infection, or other times of stress.

What mutation was detected?

The detected heterozygous variant was NM_001370658.1:c.1270G>C (p.Asp424His). This variant, NM_001370658.1:c.1270G>C (p.Asp424His), is also known as p.Asp444His or Asp446His, and is a common mutation reported in individuals with biotinidase deficiency with varying severity depending on the presence of additional BTD mutations (PubMed: [9654207](#), [21228398](#), [10206677](#)). It is important to note that the clinical spectrum of biotinidase deficiency is heterogeneous, where in some cases, patients have exhibited mild or no symptoms (PubMed: [14628140](#)). Individuals who are compound heterozygous for this mutation have presented with partial biotinidase deficiency and milder forms of the disease (PubMed: [10206677](#), [9654207](#), [12618081](#)). In the homozygous state this variant does not cause biotinidase deficiency or partial biotinidase deficiency (PubMed: [28682309](#), [9654207](#)). However, this variant in conjunction with another pathogenic variant is a common cause of partial biotinidase deficiency (PubMed: [10206677](#), [9654207](#), [12227467](#), [23644139](#)). This variant has also been observed in multiple individuals affected with profound biotinidase deficiency occurring in cis with the variant p.Ala151Thr (also known as p.Ala171Thr) and in trans with a third variant (PubMed: [10206677](#), [9654207](#)). Of note, the p.Ala151Thr variant was not observed in this individual. Individuals carrying this homozygous variant typically have 50% of mean normal serum enzyme activity (PubMed: [20539236](#), [28682309](#), [9654207](#)). The laboratory classifies this variant as pathogenic.



GENES TESTED:

Custom Beacon Preconception Carrier Screening Panel - 2 Genes

This analysis was run using the Custom Beacon Preconception Carrier Screening Panel gene list. 2 genes were tested with 100.0% of targets sequenced at >20x coverage. For more gene-specific information and assistance with residual risk calculation, see the SUPPLEMENTAL TABLE.

BTB, F2

METHODS:

Genomic DNA was isolated from the submitted specimen indicated above (if cellular material was submitted). DNA was barcoded, and enriched for the coding exons of targeted genes using hybrid capture technology. Prepared DNA libraries were then sequenced using a Next Generation Sequencing technology. Following alignment to the human genome reference sequence (assembly GRCh37), variants were detected in regions of at least 10x coverage. For this specimen, 100.00% and 100.00% of coding regions and splicing junctions of genes listed had been sequenced with coverage of at least 10x and 20x, respectively, by NGS or by Sanger sequencing. The remaining regions did not have 10x coverage, and were not evaluated. Variants were interpreted manually using locus specific databases, literature searches, and other molecular biological principles. To minimize false positive results, any variants that do not meet internal quality standards are confirmed by Sanger sequencing. Variants classified as pathogenic, likely pathogenic, or risk allele which are located in the coding regions and nearby intronic regions ($\pm$ 20bp) of the genes listed above are reported. Variants outside these intervals may be reported but are typically not guaranteed. When a single pathogenic or likely pathogenic variant is identified in a clinically relevant gene with autosomal recessive inheritance, the laboratory will attempt to ensure 100% coverage of coding sequences either through NGS or Sanger sequencing technologies ("fill-in"). All genes listed were evaluated for large deletions and/or duplications. However, single exon deletions or duplications will not be detected in this assay, nor will copy number alterations in regions of genes with significant pseudogenes. Putative deletions or duplications are analyzed using Fulgent Germline proprietary pipeline for this specimen. Bioinformatics: The FPLMv2.0 pipeline was used to analyze this specimen.

LIMITATIONS:

General Limitations

These test results and variant interpretation are based on the proper identification of the submitted specimen, accuracy of any stated familial relationships, and use of the correct human reference sequences at the queried loci. In very rare instances, errors may result due to mix-up or co-mingling of specimens. Positive results do not imply that there are no other contributors, genetic or otherwise, to future pregnancies, and negative results do not rule out the genetic risk to a pregnancy. Official gene names change over time. Fulgent uses the most up to date gene names based on HUGO Gene Nomenclature Committee (<https://www.genenames.org>) recommendations. If the gene name on report does not match that of ordered gene, please contact the laboratory and details can be provided. Result interpretation is based on the available clinical and family history information for this individual, collected published information, and Alamut annotation available at the time of reporting. This assay is not designed or validated for the detection of low-level mosaicism or somatic mutations. This assay will not detect certain types of genomic aberrations such as translocations, inversions, or repeat expansions other than specified genes. DNA alterations in regulatory regions or deep intronic regions (greater than 20bp from an exon) may not be detected by this test. Unless otherwise indicated, no additional assays have been performed to evaluate genetic changes in this specimen. There are technical limitations on the ability of DNA sequencing to detect small insertions and deletions. Our laboratory uses a sensitive detection algorithm, however these types of alterations are not detected as reliably as single nucleotide variants. Rarely, due to systematic chemical, computational, or human error, DNA variants may be missed. Although next generation sequencing technologies and our bioinformatics analysis significantly reduce the confounding contribution of pseudogene sequences or other highly-homologous sequences, sometimes these may still interfere with the technical ability of the assay to identify pathogenic alterations in both sequencing and deletion/duplication analyses. Deletion/duplication analysis can identify alterations of genomic regions which include two or more contiguous exons in size (whole blood specimens only); single exon deletions or duplications may occasionally be identified, but are not routinely detected by this test. Saliva swab samples may have reduced sensitivity for CNVs and a higher CNV failure rate. When novel DNA duplications are identified, it is not possible to discern the genomic location or orientation of the duplicated segment, hence the effect of the duplication cannot be predicted. Where deletions are detected, it is not always possible to determine whether the predicted product will remain in-frame or not. Unless otherwise indicated, deletion/duplication analysis has not been performed in regions that have been sequenced by Sanger.



Gene Specific Notes and Limitations

No gene specific limitations apply to the genes on the tested panel.

SIGNATURE:

A handwritten signature in black ink that reads "Geetu" with a stylized flourish underneath.

Geetu Mendiratta-Vij, PhD, FACMG, CGMBS on 08/17/2026
Laboratory Director, Fulgent

DISCLAIMER:

This test was developed, performed, and its performance characteristics determined by **Fulgent Therapeutics LLC** (CAP# 8042697, CLIA# 05D2043189), 4399 Santa Anita Ave., El Monte, CA 91731. It has not been cleared or approved by the FDA. The laboratory is regulated under CLIA as qualified to perform high-complexity testing. This test is used for clinical purposes. It should not be regarded as investigational or for research. Since genetic variation, as well as systematic and technical factors, can affect the accuracy of testing, the results of testing should always be interpreted in the context of clinical and familial data. For assistance with interpretation of these results, healthcare professionals may contact us directly at **(626) 350-0537** or info@fulgentgenetics.com. It is recommended that patients receive appropriate genetic counseling to explain the implications of the test result, including its residual risks, uncertainties and reproductive or medical options.

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info@fulgentgenetics.com
www.fulgentgenetics.com



To view the supplemental table describing the carrier frequencies, detection rates, and residual risks associated with the genes tested on any Beacon panel, please visit the following link:

[Beacon Expanded Carrier Screening Supplemental Table](#)





Patient Information	Specimen Information	Client Information
20055, DONOR DOB: [REDACTED] AGE: [REDACTED] Gender: M Fasting: U Phone: 725.257.0900 Patient ID: 20055	Specimen: LV589019Y Requisition: 0000042 Collected: 02/13/2025 / 09:31 PST Received: 02/14/2025 / 04:33 PST Reported: 02/24/2025 / 23:02 PST	Client #: 88807473 MAIL500 FOORD, DYLAN R LAS VEGAS SPERM BANK 2870 S MARYLAND PKWY STE 250 LAS VEGAS, NV 89109-1548

COMMENTS: FASTING:UNKNOWN

Cytogenetic Report

CHROMOSOME ANALYSIS, BLOOD - 14596
Lab:EZ
CHROMOSOME ANALYSIS, BLOOD

Order ID: 25-81138
Specimen Type: Blood
Clinical Indication: GAMETE DONOR

RESULT:
NORMAL MALE KARYOTYPE

INTERPRETATION:

Chromosome analysis revealed normal G-band patterns within the limits of standard cytogenetic analysis.

Please expect the results of any other concurrent study in a separate report.

NOMENCLATURE:

46,XY

ASSAY INFORMATION:

Method: G-Band (Digital Analysis: MetaSyst)
Cells Counted: 20
Band Level: 450
Cells Analyzed: 5
Cells Karyotyped: 3

This test does not address genetic disorders that cannot be detected by standard cytogenetic methods or rare events such as low level mosaicism or subtle rearrangements.

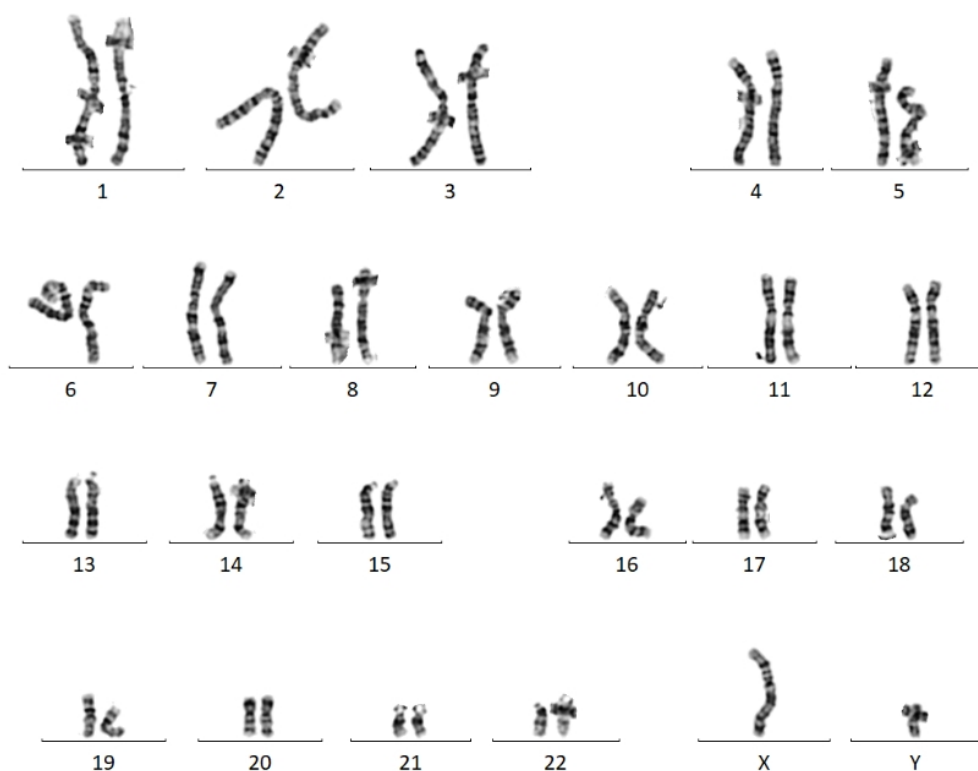
A portion of the testing was performed at SJC1

Guang Li, PhD, FACMG (800) NICHOLS-4307

Electronic Signature: 2/25/2025 1:20 AM



Patient Information	Specimen Information	Client Information
20055, DONOR DOB: [REDACTED] AGE: [REDACTED] Gender: M Fasting: U Patient ID: 20055	Specimen: LV589019Y Collected: 02/13/2025 / 09:31 PST Received: 02/14/2025 / 04:33 PST Reported: 02/24/2025 / 23:02 PST	Client #: 88807473 FOORD, DYLAN R



PERFORMING SITE:

EZ QUEST DIAGNOSTICS/NICHOLS SJC, 33608 ORTEGA HWY, SAN JUAN CAPISTRANO, CA 92675-2042 Laboratory Director: IRINA MARAMICA,MD,PHD,MBA, CLIA: 05D0643352

Date Collected: 02/13/2025

Date Received: 02/13/2025

Date Reported: 02/19/2025

Fasting: Not Given

Ordered Items: LP+12AC+CBC/D/Plt+UA+Rh+ABO...

General Comments & Additional Information

A courtesy copy of this report has been sent to 2064664696.

Date Collected: 02/13/2025

LP+12AC+CBC/D/Plt+UA+Rh+ABO...

Test	Current Result and Flag		Previous Result and Date	Units	Reference Interval
Glucose ⁰¹	97			mg/dL	70-99
Uric Acid ⁰¹	5.5			mg/dL	3.8-8.4
Therapeutic target for gout patients: <6.0					
▲ BUN ⁰¹	23	High		mg/dL	6-20
Creatinine ⁰¹	1.02			mg/dL	0.76-1.27
eGFR	97			mL/min/1.73	>59
Calcium ⁰¹	9.2			mg/dL	8.7-10.2
Protein, Total ⁰¹	6.0			g/dL	6.0-8.5
Albumin ⁰¹	4.3			g/dL	4.1-5.1
Bilirubin, Total ⁰¹	0.6			mg/dL	0.0-1.2
Alkaline Phosphatase ⁰¹	54			IU/L	44-121
LDH ⁰¹	179			IU/L	121-224
AST (SGOT) ⁰¹	23			IU/L	0-40
ALT (SGPT) ⁰¹	25			IU/L	0-44
▲ Cholesterol, Total ⁰¹	204	High		mg/dL	100-199
Triglycerides ⁰¹	62			mg/dL	0-149
HDL Cholesterol ⁰¹	60			mg/dL	>39
▲ LDL Chol Calc (NIH)	133	High		mg/dL	0-99
LDL/HDL Ratio	2.2			ratio	0.0-3.6
Please Note: ⁰¹					

LDL /HDL Ratio			
		Men	Women
1/2 Avg.Risk		1.0	1.5
Avg.Risk		3.6	3.2
2X Avg.Risk		6.2	5.0
3X Avg.Risk		8.0	6.1

Hgb Fractionation by CE: ⁰²				
Hgb F ⁰²	0.0		%	0.0-2.0
Hgb A ⁰²	97.3		%	96.4-98.8
Hgb A2 ⁰²	2.7		%	1.8-3.2
Hgb S ⁰²	0.0		%	0.0

Interpretation:⁰²

Normal hemoglobin present; no hemoglobin variant or beta thalassemia identified.
Note: Alpha thalassemia may not be detected by the Hgb Fractionation Cascade panel. If alpha thalassemia is suspected, Labcorp offers

Date Collected: 02/13/2025

LP+12AC+CBC/D/Plt+UA+Rh+ABO... (Cont.)

Alpha-Thalassemia DNA Analysis (#511172).

ABO Grouping ⁰¹	B		
Rh Factor ⁰¹	Positive		
	Please note: Prior records for this patient's ABO / Rh type are not available for additional verification.		
. ⁰¹			
CBC, Platelet Ct, and Diff ⁰¹			
WBC ⁰¹	7.2	x10E3/uL	3.4-10.8
RBC ⁰¹	5.02	x10E6/uL	4.14-5.80
Hemoglobin ⁰¹	15.3	g/dL	13.0-17.7
Hematocrit ⁰¹	44.8	%	37.5-51.0
MCV ⁰¹	89	fL	79-97
MCH ⁰¹	30.5	pg	26.6-33.0
MCHC ⁰¹	34.2	g/dL	31.5-35.7
RDW ⁰¹	12.3	%	11.6-15.4
Platelets ⁰¹	344	x10E3/uL	150-450
Neutrophils ⁰¹	60	%	Not Estab.
Lymphs ⁰¹	25	%	Not Estab.
Monocytes ⁰¹	9	%	Not Estab.
Eos ⁰¹	5	%	Not Estab.
Basos ⁰¹	1	%	Not Estab.
Neutrophils (Absolute) ⁰¹	4.3	x10E3/uL	1.4-7.0
Lymphs (Absolute) ⁰¹	1.8	x10E3/uL	0.7-3.1
Monocytes(Absolute) ⁰¹	0.6	x10E3/uL	0.1-0.9
Eos (Absolute) ⁰¹	0.3	x10E3/uL	0.0-0.4
Baso (Absolute) ⁰¹	0.1	x10E3/uL	0.0-0.2
Immature Granulocytes ⁰¹	0	%	Not Estab.
Immature Grans (Abs) ⁰¹	0.0	x10E3/uL	0.0-0.1
. ⁰¹			
Urinalysis Gross Exam ⁰¹			
Specific Gravity ⁰¹	1.017		1.005-1.030
pH ⁰¹	7.0		5.0-7.5
Urine-Color ⁰¹	Yellow		Yellow
Appearance ⁰¹	Clear		Clear
► WBC Esterase ⁰¹	1+	Abnormal	Negative
Protein ⁰¹	Negative		Negative/Trace
Glucose ⁰¹	Negative		Negative
Ketones ⁰¹	Negative		Negative
Occult Blood ⁰¹	Negative		Negative
Bilirubin ⁰¹	Negative		Negative
Urobilinogen,Semi-Qn ⁰¹	0.2	mg/dL	0.2-1.0
Nitrite, Urine ⁰¹	Negative		Negative
Microscopic Examination ⁰¹	See below: Microscopic was indicated and was performed.		

Date Collected: 02/13/2025

LP+12AC+CBC/D/Plt+UA+Rh+ABO... (Cont.)

WBC ⁰¹	0-5	/hpf	0-5
RBC ⁰¹	None seen	/hpf	0-2
Epithelial Cells (non renal) ⁰¹	None seen	/hpf	0-10
Casts ⁰¹	None seen	/lpf	None seen
Bacteria ⁰¹	None seen		None seen/Few

Disclaimer
The Previous Result is listed for the most recent test performed by Labcorp in the past 5 years where there is sufficient patient demographic data to match the result to the patient. Results from certain tests are excluded from the Previous Result display.

Icon Legend
▲ Out of Reference Range ■ Critical or Alert

Performing Labs
01: PDLCA - Labcorp Phoenix, 5005 S 40th Street Ste 1200, Phoenix, AZ 85040-2969 Dir: Earle Collum, MD
02: BN - Labcorp Burlington, 1447 York Court, Burlington, NC 27215-3361 Dir: Sanjai Nagendra, MD
For inquiries, the physician may contact Branch: 888-522-2677 Lab: 800-788-9743

Patient Details 20055, Donor Phone: Date of Birth: [REDACTED] Age: [REDACTED] Sex: Male Patient ID: 20055 Alternate Patient ID: 20055	Physician Details D FOORD Las Vegas Sperm Bank 2870 S Maryland Pkwy Ste 250, Las Vegas, NV, 89109 Phone: 206-588-1484 Account Number: 27021640 Physician ID: NPI: 1063683969	Specimen Details Specimen ID: 044-592-0365-0 Control ID: L2501180929 Alternate Control Number: L2501180929 Date Collected: 02/13/2025 0929 Local Date Received: 02/13/2025 0000 ET Date Entered: 02/13/2025 2034 ET Date Reported: 02/19/2025 0608 ET
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